



Docket No.: 31126/41458UTL  
(PATENT)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of: Ledbetter *et al.*

Application No.: 10/053,530

Confirmation No.: 8993

Filed: January 17, 2002

Art Unit: 1642

For: Binding Domain Immunoglobulin Fusion Proteins

Examiner: Blanchard, David

**DECLARATION OF ALAN F. WAHL**

1. I, Alan F. Wahl, declare as follows.

2. I am the Vice President, Technology Research, at Trubion Pharmaceuticals, Inc., the assignee of the above-identified patent application. A copy of my Curriculum Vitae is attached hereto as Exhibit 1.

3. Various single chain proteins were made that include a binding domain capable of binding a target biological molecule joined to a IgG hinge peptide with two cysteine residues (one at the cysteine position responsible for forming a disulfide bond with a light chain constant region in a naturally occurring IgG antibody) further joined to immunoglobulin heavy chain CH2 and CH3 constant region polypeptides. A number of the single chain proteins were demonstrated to promote antibody-dependent, cell-mediated cytotoxicity (ADCC) and/or complement fixation.

4. Examples of such single chain proteins that were made are:

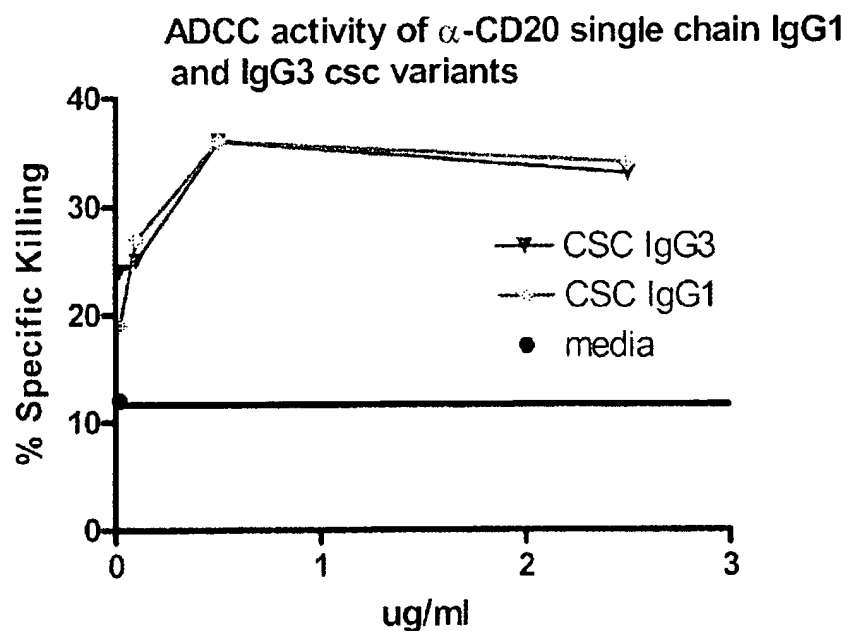
Single chain protein	Binding domain target	Hinge	CH2	CH3
1 (TRU-015)	CD20	IgG <sub>1</sub> CSC	IgG <sub>1</sub>	IgG <sub>1</sub>
2	CD20	IgG <sub>1</sub> CCS	IgG <sub>1</sub>	IgG <sub>1</sub>
3	CD37	IgG <sub>1</sub> CSC	IgG <sub>1</sub>	IgG <sub>1</sub>
4	CD20	IgG <sub>3</sub> CSC	IgG <sub>1</sub>	IgG <sub>1</sub>

5. Single chain proteins 1 and 4 were tested and found to promote ADCC and

complement fixation.

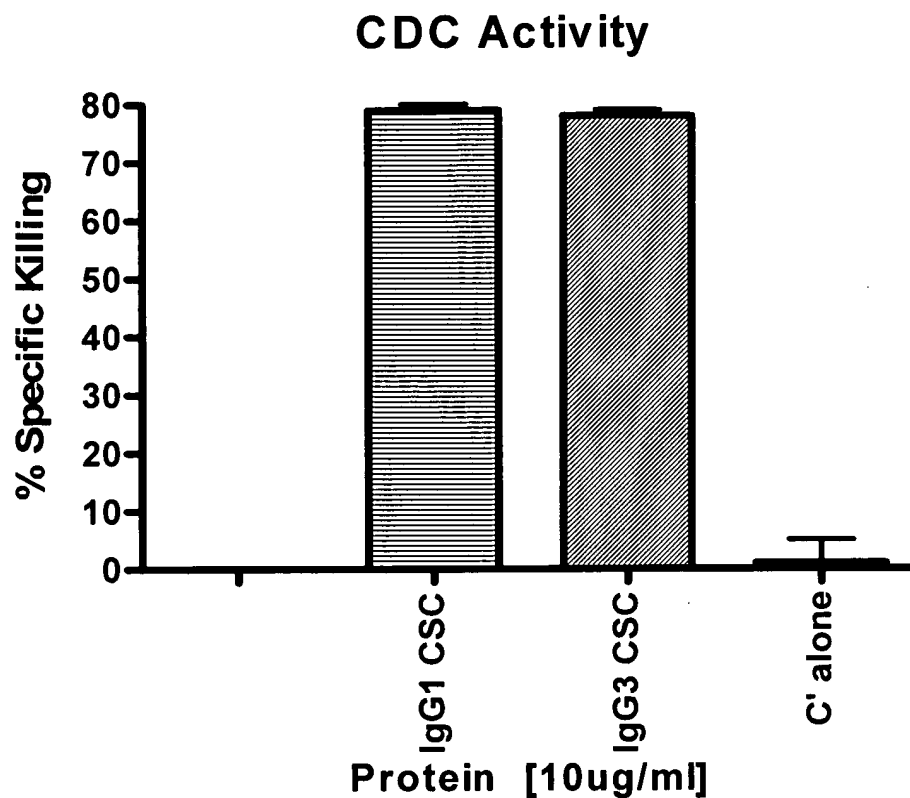
6. The ADCC assays were conducted as follows. Briefly,  $1 \times 10^7$ /ml BJAB B-cells were labeled with 500  $\mu\text{Ci}/\text{ml}$   $^{51}\text{Cr}$  sodium chromate (#CJS1, Amersham Biosciences, Piscataway, NJ) for 2 hrs at  $37^\circ\text{C}$  in Iscoves media (#12440-053, Gibco/Invitrogen, Grand Island, NY) with 10% FBS (#16140-071, Gibco/Invitrogen, Grand Island, NY). The  $^{51}\text{Cr}$  loaded BJAB were then washed 3 times in RPMI (#11875-093, Gibco/Invitrogen, Grand Island, NY) media with 10% FBS and resuspended at  $4 \times 10^5$ /ml in RPMI. Peripheral blood mononuclear cells (PBMC) from in-house donors were isolated from heparinized whole blood via centrifugation over Lymphocyte Separation Medium (#50494, MP Biomedicals, Aurora, OH), washed 2 times with RPMI media and resuspended at  $5 \times 10^6$ /ml in RPMI with 10% FBS. Single chain proteins were added to RPMI media with 10% FBS at 4 times the final concentration and three 10-fold serial dilutions for each single chain protein were prepared. These single chain proteins were then added to 96-well U- bottom plates at 50  $\mu\text{l}/\text{well}$  for the indicated final concentrations. The  $^{51}\text{Cr}$  labeled BJAB were then added to the plates at 50  $\mu\text{l}/\text{well}$  ( $2 \times 10^4$ /well). The PBMC were then added to the plates at 100  $\mu\text{l}/\text{well}$  ( $5 \times 10^5$ /well) for a final ratio of 25:1 effectors (PBMC):target (BJAB). Effectors and targets were added to media alone to measure background killing. The  $^{51}\text{Cr}$  labeled BJAB were added to media alone to measure spontaneous release of  $^{51}\text{Cr}$  and to media with 5% NP40 (#28324, Pierce, Rockford, IL) to measure maximal release of  $^{51}\text{Cr}$ . The plates were incubated for 6 hours at  $37^\circ\text{C}$  in 5% $\text{CO}_2$ . Fifty  $\mu\text{l}$  (can use 25  $\mu\text{l}$ ) of the supernatant from each well were then transferred to a LumaPlate-96 (#6006633, Perkin Elmer, Boston, MA) and dried overnight at room temperature. In the morning, cpm were read on a Packard TopCount-NXT. Percent specific killing was calculated by subtracting cpm (mean of quadruplicate samples) of sample – cpm spontaneous release/cpm maximal release-cpm spontaneous release X 100. Figure 1 below shows the results of the assays.

Figure 1



7. The complement fixation assays were conducted as follows. In brief, approximately  $3 \times 10^5$  Ramos B-cells were added per well to 96-well V-bottomed plates in 50 ul of Iscoves (#12440-053, Gibco/Invitrogen, Grand Island, NY) media (no FBS). The single chain proteins in Iscoves, (or Iscoves alone) were added to the wells in 50 ul at 2 times the indicated final concentration. The cells and single chain proteins were incubated for 45 minutes at 37°C. The cells were washed 2 ½ times in Iscoves with no FBS and resuspended in Iscoves with human serum (# A113, Quidel, San Diego, CA) in the 96-well plate at the indicated concentrations. The cells were then incubated for 90 minutes at 37°C. The cells were washed by centrifugation and resuspended in 125 ul cold PBS. The cells were transferred to FACs cluster tubes (#4410, CoStar, Corning, NY) and 125 ul PBS with propidium iodide (# P-16063, Molecular Probes, Eugene, OR) at 5 ug/ml was added. The cells were incubated with the propidium iodide for 15 minutes at room temperature in the dark and then placed on ice and read and analyzed on a FACsCalibur with CellQuest software (Becton Dickinson). The results of the assays are shown in Figure 2 below.

Figure 2



8. Single chain protein 1 (TRU-015) also exhibited a clinical benefit for rheumatoid arthritis patients in a Phase IIa study in humans. Results of the study were presented at the American College of Rheumatology Annual Scientific Meeting, Washington, DC during the week of November 10-15, 2006. A copy of the poster presented is attached as Exhibit 3. Furthermore, enrollment and dosing of patients has been completed for a Phase IIb clinical trial in human rheumatoid arthritis patients.

9. Other single chain proteins, described below, were made that include a binding domain capable of binding a target biological molecule joined to an IgG hinge peptide with one cysteine residue (the one cysteine being at the position responsible for forming a disulfide bond with a light chain constant region in a naturally occurring IgG antibody) further joined to immunoglobulin heavy chain CH2 and CH3 constant region polypeptides.

Single chain protein	Binding domain target	Hinge	CH2	CH3
5	CD20	IgG <sub>3</sub> CSS	IgG <sub>1</sub>	IgG <sub>1</sub>
6	CD20	IgG <sub>1</sub> CSS	IgG <sub>1</sub>	IgG <sub>1</sub>
7	CD37	IgG <sub>1</sub> CSS	IgG <sub>1</sub>	IgG <sub>1</sub>

10. Like the two-cysteine single chain proteins, single chain protein 5 promoted ADCC and complement fixation activities when tested in the assays described above. See Figures 3 and 4.

Figure 3

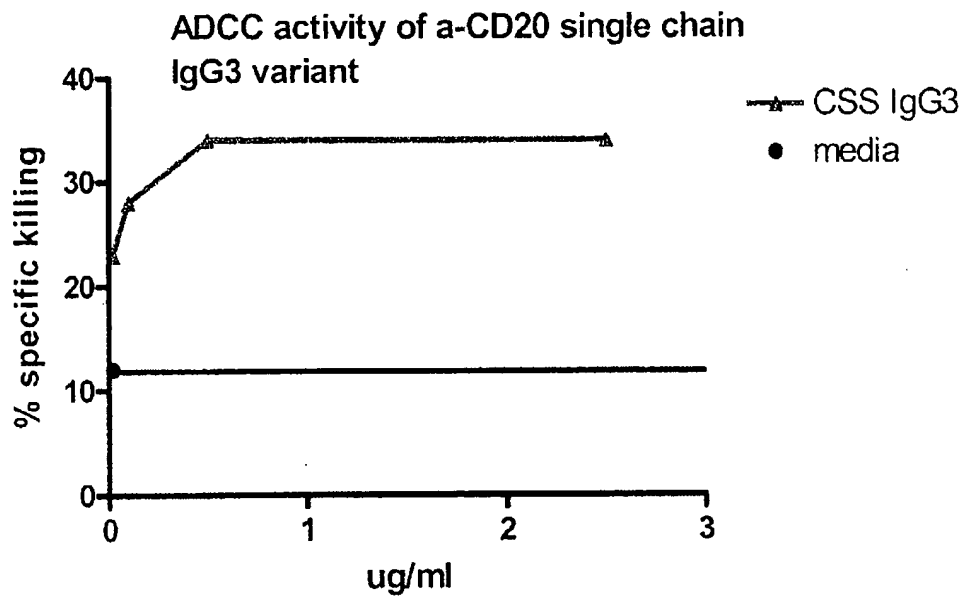
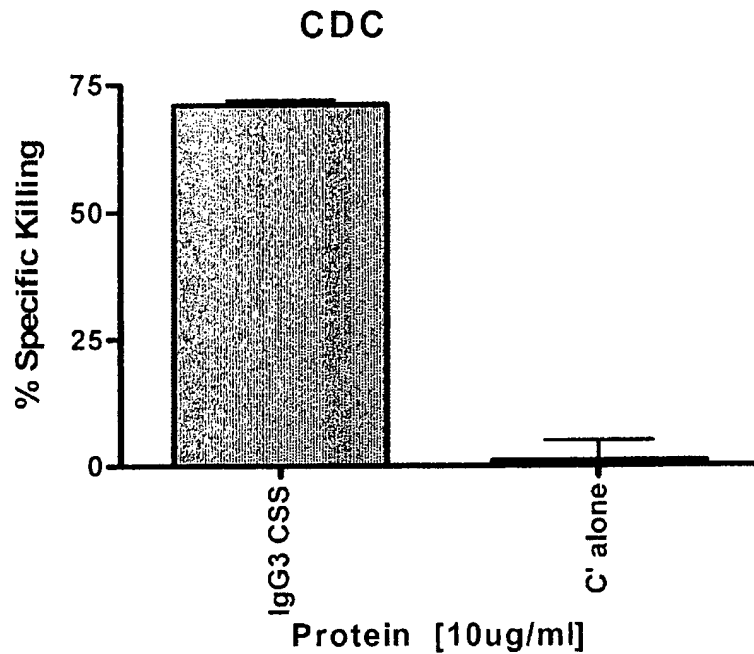


Figure 4



11. I declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the above-identified application or any patent issuing thereon.

3/20/02  
Date

Alan F. Wahl

# Exhibit 1

## CURRICULUM VITAE

NAME: Alan Francis Wahl

ADDRESS: 6150 East Mercer Way  
Mercer Island WA, 98040  
(206) 232-2783 (h)  
(425) 527-4610 (w)  
email: [westwahl@comcast.net](mailto:westwahl@comcast.net)

INSTITUTION	DEGREE	YEAR
University of Rochester Rochester, NY	Ph.D. Biochemistry & Biophysics	1986
University of Rochester Rochester, NY	M.S. Biochemistry & Biophysics	1984
Rochester Institute of Technology, Rochester, NY	B.S. Biology/Chemistry <i>cum laude</i>	1978
Stanford University Stanford, CA	Postdoctoral Fellowship Experimental Oncology	1986-1989

### PROFESSIONAL EXPERIENCE:

**2006-present** Vice President, Technology Research  
Trubion Pharmaceuticals, Inc. Seattle, WA 98121

#### Areas of Scientific Excellence:

- Discovery of novel biological immunotherapeutics for treatment of cancer and immunologic diseases.

#### Management Expertise:

- Member of Sr. management team responsible for corporate technical and strategic decisions.
- Scientific liaison for multiple strategic partners in Immunology and Oncology.
- Direct responsibility for Department of Technology Research of 14+ researchers

**2000-2006** Senior Director, Molecular Oncology and Immunology  
Seattle Genetics, Inc., Bothell, WA 98021

#### Areas of Scientific Excellence:

- Discovery and preclinical development of targeted biologics and small molecules for treatment of cancer and immunologic diseases.
- Identified and advanced numerous preclinical candidates
- Led preclinical IND teams for all Seattle Genetics' drugs currently in development or in clinical trials.
- Responsible for a strong in-house discovery/ development pipeline.

#### Management Expertise:

- Founding Director, Departments of Molecular Oncology and Immunology.
- Instrumental in building Seattle Genetics, Inc. from its 5-person inception to currently 140+ employees.
- Member of Senior Management Team reporting to CSO and Sr. VP.

- Chair, Late Stage Project Review Committee includes all aspects of drug discovery and experimental therapeutics.
- Key company representative for numerous in-licensing, out-licensing, investment and strategic alliances.
- Scientific liaison for multiple strategic partners in Immunology and Oncology.
- Direct responsibility for Departments of Molecular Oncology and Immunology and co-responsibility for Experimental Therapeutics Department. The former group includes an Associate Director and 12 researchers; the latter group includes 8 researchers.

**1998-2000 Director, Biochemistry and Tumor Biology**  
**Seattle Genetics, Inc., Bothell, WA 98021**

Management Areas: Established company core competencies in Immunology, Tumor Cell Biology, and Experimental Therapeutics for drug discovery and development. Directly organized, staffed and grew departments of biology drug discovery and experimental therapeutics. Helped staff and shape Clinical, Development, Chemistry and Preclinical Development Departments for the newly formed company. Lead preclinical discovery and development teams, and successfully represented their accomplishments, including IND enabling studies to the company, investors and collaborators.

**1997-1998 Principal Scientist**  
**Department of Immunology**  
**Zymogenetics Inc. Seattle WA 98102**

Research Areas: Discovery and early stage development of biomolecules for treating immunologic diseases. Established immunologic / biochemical and tumor cell-based screens to detect signal transduction, proliferation and activation control of leukocytes and tumor cells.

**1996-97 Principal Scientist and Group Leader**  
**1992-1996 Senior Research Investigator II**  
**Department of Inflammation, Bristol-Myers Squibb Pharmaceutical Research Institute,**  
**Seattle WA, 98121**

Research Areas: Mechanisms of apoptosis, proliferation and cell cycle control of leukocytes and neoplastic cells. Small molecule and biomolecule drug discovery for treatment of cancer and autoimmune diseases. Lead Scientist for development of proprietary cytokine clinical candidate for treatment of inflammatory diseases.

Management Areas: Responsible for evaluations and review of research of seven staff scientists and associates and for management review of external proposals to corporation. Participated in site visits for potential company or technology acquisitions. Direct reporting to V.P. Inflammation and Immunology. Responsible for departmental reporting to Bristol-Myers Squibb Corporate Drug Discovery Management Team.

**1991-1992 Senior Research Investigator, Antitumor Cellular and Molecular Biology.**  
**1989-1991 Research Investigator**

**Bristol-Myers Squibb Pharmaceutical Research Institute, Wallingford, CT**

Research Areas: Mechanism of anti-cancer drug action, platinol and taxane analog programs. Contributing scientist to TAXOL and Carboplatin programs. Enzymology of gene amplification and mechanisms of drug resistance in neoplasia.

Management Areas: Directed Core Facility for Synthetic Biomolecules, Bristol-Myers Squibb PRI, Wallingford, CT

**POSTDOCTORAL RESEARCH FELLOWSHIP**

**1986-1989**

**Stanford University School of Medicine, Stanford, CA**  
**Laboratory of Experimental Oncology, Department of Pathology**  
**Advisors: Drs. T.S.-F. Wang and D. Korn**



Research Area: Protein/DNA interactions during DNA replication in normal and neoplastic cell proliferation. Cloning, transcriptional regulation and cell cycle analysis of human DNA polymerase  $\alpha$ .

## **GRADUATE RESEARCH**

1981-1986

University of Rochester, Rochester, NY

Department of Biochemistry & Biophysics and The Cancer Center

Thesis Advisors: Drs. R.A. Bambara and E.M. Lord

Research Area: Purification, physicochemical characterization and enzymology of eukaryotic DNA dependent ATPases and replicative DNA polymerases. Cell fusion and monoclonal antibody production.

Thesis: *Mechanisms and Control of Mammalian DNA Replication.*

**Awards:** National Institutes of Health Molecular Genetics Predoctoral Fellowship 1981-1985  
Stanford University Deans Fellowship 1986-1987  
National Cancer Institute Tumor Biology Training Fellowship 1987-1989

### **Society & Committee Memberships:**

American Association for Cancer Research

American Association for the Advancement of Science

American Heart Association Review Committee 1994- 2000.

Member, Councils of Advisors, Gerson Lehrman Group, New York, NY

Ad hoc Reviewer for *Journal of Biological Chemistry*, *Biochemistry*, *Cancer Research*, *Clinical Cancer Research*, *Molecular and Cellular Biology*, *Blood*, *Bioconjugate Chemistry* and *Nature Medicine*.

### **STUDENT MENTORING:**

Russell Sanderson	Ph.D. Postdoctoral Fellow	2003-2004
Andrew McShea	Ph.D. Postdoctoral Fellow	1996-1997
Mary Horne	Ph.D. Postdoctoral Fellow	1993-1995
Ellen Carmichael	Ph.D. Postdoctoral Fellow	1990-1992
Brian Spain	M.S. Stanford University	1987-1988

### **RESEARCH / RESEARCH MANAGEMENT SKILLS:**

**Molecular Biology:** All stages of cDNA cloning and sequencing, PCR, Northern and Southern analysis, transcriptional regulation, analysis, and gene expression.

**Cell Biology:** Mammalian tissue culture, cell fusion, pathogenic microbiology, immunocytochemistry, in situ transcription, cell cycle separation and FACS analysis.

**Tumor Biology:** Tumor and primary cell culture, drug/drug interaction, growth control and apoptosis. Design, development and evaluation of in xenograft efficacy models of human carcinoma and hematologic neoplasia.

**Inflammation:** Design, development and evaluation of efficacy studies *in vitro* and animal models of arthritis, inflammatory bowel disease, pulmonary inflammation and multiple sclerosis.

**Protein Chemistry:** Purification: HPLC, FPLC, RP, ion exchange, gel permeation and ligand affinity chromatography; chromato- and isoelectric focusing; synthesis of protein and nucleic acid based matrices, chromogenic and isotopic labeling.

**Analysis:** Peptide sequencing, Amino Acid Analysis, PAGE, thin layer, 2-D peptide mapping, rate sedimentation and kinetics. ABI advanced peptide sequencer training.

**Drug Development:** Authored preclinical sections to INDs for acytokine anti-inflammatory, a small molecule anti-cancer agent, and multiple antibody and antibody-drug conjugate anti-cancer clinical candidates. Authored numerous SOPs for drug development candidates.

**Management:** Founded, developed and currently successfully manage two, highly productive departments within the organization. Organized and chaired committee for review and prioritization of late stage projects and represented these projects to corporate portfolio review committee.

#### **PREDOCTORAL EMPLOYMENT**

**1979-1981** Senior Research Associate, Biochemistry Division, Eastman Kodak Research, Rochester, NY. Development of solid-state immunochemical assays. Purification of serum proteins, antigen/antibody production, biomolecule immobilization on synthetic supports.

**1978** Research Technician, U.S. Centers for Disease Control, Albany, NY. Developed immunodiagnosics for Legionnaire's disease/Rocky mountain spotted fever. Rickettsia cell surface protein isolation and development of solid-phase immunoassays.

**1976-1977** Area Administrator, National Technical Institute for the Deaf/R.I.T., Rochester NY. While a full time undergraduate student, directed a staff of 35 resident advisors to oversee a 2000-person residence complex for the hearing impaired.

#### **TEACHING EXPERIENCE**

**1990** Adjunct Instructor, Department of Biological Sciences, Wesleyan University, Middletown, CT. Instructed graduate course in Oncogenes and Growth Factors.

**1982-1983** Teaching Assistant, Department of Biochemistry, University of Rochester Medical Center, Rochester NY. Graduate courses in Biochemistry and Advanced Biochemistry.

**1976-1978** Instructor, National Technical Institute for the Deaf/ R.I.T. Co-developed curriculum and instructed leadership training and experiential education courses.

## PUBLICATIONS

1. **A.F. Wahl**, J.W. Hockensmith, S.P. Kowalski and R.A. Bambara. (1983) Alternative explanation for excision repair deficiency caused by the polAexI mutation. *J. Bacteriology*, 155(2):982-986.
2. **A.F. Wahl**, S.P. Kowalski, L.W. Harwell, E.M. Lord, and R.A. Bambara. (1984) Immunoaffinity purification and properties of a high molecular weight calf thymus DNA-alpha polymerase. *Biochemistry* 23(9):1895-1899.
3. R.A. Bambara, J.J. Crute and **A.F. Wahl**. (1985) Is Ap4A an activator of eukaryotic DNA replication? *Cancer Investigation*. 3:473-479.
4. J.J. Crute, **A.F. Wahl**, R.A. Bambara. (1986) Purification and characterization of DNA polymerase delta from calf thymus. *Biochemistry* 25:26-36.
5. J.J. Crute, **A.F. Wahl**, R. Murant, R.A. Bambara, S.L. Gibson and R. Hilf. (1986) Inhibition of DNA polymerases in vitro and in vivo by hematoporphyrin derivative and photoradiation. *Cancer Research*. 46:153-159.
6. J.W. Hockensmith, **A.F. Wahl**, S. Kowalski and R.A. Bambara. (1986) Purification of calf thymus DNA-dependent ATPase that prefers a primer-template junction effector. *Biochemistry* 25:7812-7821.
7. **A.F. Wahl**, J.J. Crute, R.D. Sabatino, J.B. Bodner, R.L. Marraccino, L.W. Harwell, E.M. Lord and R.A. Bambara. (1986) Properties of two forms of DNA polymerase delta from calf thymus. *Biochemistry* 25: 7821-7827.
8. R.L. Marraccino, **A.F. Wahl**, P.C. Keng, E.M. Lord and R.A. Bambara. (1987) Cell cycle dependent activities of DNA polymerase alpha and delta in Chinese hamster ovary cells. *Biochemistry* 26:7864-7870.
9. S.W. Wong, **A.F. Wahl**, P.M. Yuan, N. Arai, B.E. Pearson, K. Arai, D. Korn, M. Hunkapiller and T.S.-F. Wang. (1988) Human DNA polymerase  $\alpha$  gene expression is cell proliferation dependent and its primary structure is similar to both prokaryotic and eukaryotic replicative DNA polymerases. *EMBO J.* 7:37-47.
10. **A.F. Wahl**, A. Moore Geis, Brian H. Spain, W.W. Wong, D. Korn and T.S.-F. Wang. (1988) Gene expression of human DNA polymerase  $\alpha$  during cell proliferation and the cell cycle. *Molecular and Cellular Biology* 8:5016-5025.
11. H.P. Nasheuer, A. Moore, **A.F. Wahl** and T.S.-F. Wang. (1991) Cell cycle-dependant phosphorylation of human DNA polymerase  $\alpha$ . *J. Biol. Chem.* 266:7893-7903.

12. E.P. Carmichael, J. Roome and **A.F. Wahl**. (1993) Binding of a sequence-specific single stranded DNA binding factor to the SV40 core origin IR domain is cell cycle regulated. *Molecular and Cellular Biology* 13: 408-420.
13. L. Huang, J. J. Turchi, **A.F. Wahl** and R. A. Bambara. (1993) Effects of the anticancer drug *cis*-diamminedichloroplatinum (II) on the activities of calf thymus DNA polymerase  $\epsilon$ . *Biochemistry* 32: 841-848.
14. L. Huang, J. J. Turchi, **A. F. Wahl** and R. A. Bambara. (1993) Activity of Calf Thymus DNA helicase  $\epsilon$  on *cis*- diamminedichloroplatinum (II) damaged DNA. *J. Biol. Chem.* 268: 26731-26737.
15. K. Donaldson, G. Goolsby and **A. F. Wahl**. (1994) Cytotoxicity of the anticancer agents cisplatin and taxol during cell proliferation and the cell cycle. *Int. J. Cancer* 57: 847-855.
16. K. Donaldson, G. Goolsby, P. A. Kiener and **A. F. Wahl**. (1994) Activation of p34<sup>cdc2</sup> coincident with taxol-induced apoptosis. *Cell Growth and Differentiation* 5: 1041-1050.
17. G. L. Schieven, **A. F. Wahl**, S. Myrdal, L. Grosmaire and J. A. Ledbetter (1995) Lineage specific induction of B cell apoptosis and altered signal transduction by the phosphotyrosine phosphatase inhibitor Bis(maltolato) oxovanadium(IV). *J. Biol. Chem.* 270: 20824-20831.
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20. M. C. Horne, G. L. Goolsby, K. L. Donaldson, D. Tran, M. Nuebauer and **A. F. Wahl** (1996) Cyclin G1 and G2 comprise a new family of cyclins with contrasting tissue specific and cell cycle regulated expression. *J. Biol. Chem.* 271:6050-6061.
21. N. Malik, B. W. Greenfield, **A. F. Wahl** and P. A. Kiener (1996) Activation of human monocytes through CD40 induces matrix metalloproteinases. *J. Immunol.* 156:3952-3960.
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24. M. C. Horne, K. L. Donaldson, G. L. Goolsby, D. Tran, M. Mulheisen, J. W. Hell, and **A. F. Wahl** (1997) Cyclin G2 is upregulated during growth inhibition and B cell antigen receptor-mediated cell cycle arrest. *J. Biol. Chem.* 272:12650-12661.
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31. **A. F. Wahl** and K. L. Donaldson (1999) Separation of cells into progressive stages of the cell division cycle by counterflow centrifugal elutriation. *Current Protocols in Cell Biology*, Unit 8.6. (J. Wiley and Sons, New York, NY).
32. M. Jung, **A. F. Wahl**, W. Neupert, G. Geisslinger, and P. D. Senter (2000) Synthesis and activities of fluorinated derivatives of sulindac sulfide and sulindac sulfone. *Pharm. Pharmacol.* 6:217-221.
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34. **A. F. Wahl**, K. L. Donaldson, B. J. Mixan, P. A. Trail and C. B. Siegall (2001) Selective Tumor Sensitization to Taxanes with mAb-Drug Conjugate cBR96-Doxorubicin. *International Journal of Cancer*. 93:590-600.
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37. B.E. Toki, C. G. Cervený, **A. F. Wahl**, and P.D. Senter (2002) Protease mediated fragmentation of p-amidobenzylethers: A new strategy for the activation of anticancer agents. *Journal of Organic Chemistry*, 67:1866-1872.
38. **A. F. Wahl**, K. Klussman, J. Thompson, G. Risdon, D. Chace, C. B. Siegall and J. A. Francisco (2002) The anti-CD30 mAb SGN-30 induces apoptosis and affects antitumor activity in models of Hodgkin's Disease. *Cancer Research*, 62: 3736-3742.
39. S. O. Doronina, B. E. Toki, B. Mendelsohn, M. Torgov, C. G. Cervený, D. F. Chace, S. Rejniak, R. DeBlanc, R. P. Gearing, C. B. Siegall, J. A. Francisco, **A. F. Wahl**, D. L. Myer, and D. D. Senter (2003), Development of potent monoclonal antibody auristatin conjugates for cancer therapy. *Nature Biotechnology*, 21: 778-794.
40. J. A. Francisco, C. G. Cervený, D. L. Meyer, B. J. Mixan, K. Klussman, D. F. Chace, S. Rejniak, K. Gordon, R. DeBlanc, B. E. Toki, C.-L. Law, S. O. Doronina, C. B. Siegall, P. D. Senter and **A. F. Wahl** (2003) SGN-35, an anti-CD30-monomethyl auristatin E conjugate with potent and selective antitumor activity. *Blood*, 102: 1458-1465.
41. K. Klussman, B. Mixan, D. L. Meyer, P. D. Senter and **A. F. Wahl** (2004) Secondary mAb-vcMMAE conjugates are highly sensitive reporters of antibody internalization *via* the lysosome pathway. *Bioconjugate Chemistry*, 15:765-773.
42. J. Petroziello, A. Yamane, L. Westendorf, M. Thompson, C. McDonagh, C. Cervený, C.-L. Law, **A. F. Wahl** and P. Carter, (2004) Suppression subtractive hybridization and expression profiling identifies a unique set of genes over-expressed in non-small cell lung cancer. *Oncogene*, 23:7734-7745.
43. C.-L. Law, C. G. Cervený, K. Klussman, D. F. Chace, K. A. Gordon, D. L. Meyer, S. O. Doronina, C. B. Siegall, P. D. Senter, J. A. Francisco and **A. F. Wahl**. (2004). Efficient elimination of B-lineage lymphomas by anti-CD20 auristatin conjugates. *Clinical Cancer Research*, 10:7842-7851. (*Feature /Cover Article*)
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SGN-30. A chimeric monoclonal antibody for the treatment of hematologic malignancies. Seattle Genetics, Inc.  
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Exhibit 2

BEST AVAILABLE COPY

# TRU-015, A SMALL MODULAR IMMUNOPHARMACEUTICAL (SMIP™) DRUG CANDIDATE DIRECTED AGAINST CD20, DEMONSTRATES CLINICAL IMPROVEMENT IN SUBJECTS WITH RHEUMATOID ARTHRITIS

D. J. Burge,<sup>1</sup> C. Shu,<sup>2</sup> R. W. Martin,<sup>3</sup> T. W. Littlejohn,<sup>4</sup> D. J. Wallace,<sup>5</sup> J. Taborn,<sup>6</sup> W. R. Palmer,<sup>7</sup> A. Kivitz<sup>8</sup>

<sup>1</sup>Trubion Pharm., Seattle, WA; <sup>2</sup>Wyeth, Collegeville, PA; <sup>3</sup>Arth Ed & Treatment Ctr, Grand Rapids, MI; <sup>4</sup>Piedmont Med Res Assoc, Winston-Salem, NC; <sup>5</sup>Wallace Rheum Study Ctr, Los Angeles, CA; <sup>6</sup>Midwest Arth Ctr, Kalamazoo, MI; <sup>7</sup>Westroads Med Gp, Omaha, NE; <sup>8</sup>Altosna Ctr for Clin Res, Duncansville, PA

## Abstract

**BACKGROUND:** Protein therapeutics directed against CD20-antigen on B lymphocytes have been demonstrated to be highly effective in the treatment of rheumatoid arthritis (RA). However, these therapies are associated with a number of side effects, including the development of anti-drug antibodies, which may limit their efficacy. TRU-015 is a small modular immunopharmaceutical (SMIP) drug candidate that selectively targets B lymphocytes in RA, but does not bind to CD20-antigen on other cells. TRU-015 is a CD20-directed therapy, a dose-dependent disease-modifying antirheumatic drug (DMARD) that is designed to improve the efficacy of RA therapies while minimizing the side effects associated with current therapies.

**OBJECTIVE:** The purpose of this study was to evaluate the safety and efficacy of TRU-015 in subjects with RA. The study was designed to evaluate the safety and efficacy of TRU-015 in subjects with RA, and to determine the optimal dose of TRU-015.

**DESIGN:** This was a Phase 1 study, a dose-escalation study in subjects with RA. The study was designed to evaluate the safety and efficacy of TRU-015 in subjects with RA, and to determine the optimal dose of TRU-015. The study was a Phase 1 study, a dose-escalation study in subjects with RA. The study was designed to evaluate the safety and efficacy of TRU-015 in subjects with RA, and to determine the optimal dose of TRU-015.

**SETTING:** The study was conducted at several sites, including the University of Washington Medical Center, the University of California, San Francisco, and the University of Texas at Austin.

**PARTICIPANTS:** The study included 10 subjects with RA, who were enrolled in the study after giving informed consent.

**INTERVENTIONS:** The study included a placebo group and a group receiving TRU-015. The TRU-015 group received a single intravenous infusion of TRU-015 at a dose of 10 mg/kg.

**MEASUREMENTS AND MAIN RESULTS:** The study found that TRU-015 was well-tolerated and that it significantly improved the clinical response in subjects with RA. The study also found that TRU-015 was effective in reducing the levels of anti-drug antibodies.

**CONCLUSIONS:** The study found that TRU-015 was well-tolerated and that it significantly improved the clinical response in subjects with RA. The study also found that TRU-015 was effective in reducing the levels of anti-drug antibodies.

## Background

**Small Modular Immunopharmaceutical (SMIP) drugs**

SMIP drugs are a new class of immunopharmaceuticals that are designed to target specific cells or molecules in the body. They are composed of a small protein that is fused to a therapeutic agent. This design allows them to be highly specific and to have a reduced risk of side effects compared to traditional biologics.

TRU-015 is a SMIP drug candidate that is designed to target B lymphocytes in RA. It is a CD20-directed therapy, which means it targets the CD20 protein on the surface of B lymphocytes. This protein is a key component of the B cell receptor, and its expression is increased in B lymphocytes in RA. By targeting CD20, TRU-015 is designed to deplete B lymphocytes and reduce the inflammation associated with RA.

TRU-015 is a small modular immunopharmaceutical (SMIP) drug candidate that is designed to target B lymphocytes in RA. It is a CD20-directed therapy, which means it targets the CD20 protein on the surface of B lymphocytes. This protein is a key component of the B cell receptor, and its expression is increased in B lymphocytes in RA. By targeting CD20, TRU-015 is designed to deplete B lymphocytes and reduce the inflammation associated with RA.

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## Methods

**Study Design**

The study was a Phase 1 study, a dose-escalation study in subjects with RA. The study was designed to evaluate the safety and efficacy of TRU-015 in subjects with RA, and to determine the optimal dose of TRU-015. The study was a Phase 1 study, a dose-escalation study in subjects with RA. The study was designed to evaluate the safety and efficacy of TRU-015 in subjects with RA, and to determine the optimal dose of TRU-015.

**Subjects**

The study included 10 subjects with RA, who were enrolled in the study after giving informed consent. The subjects were selected based on the following criteria: they had a confirmed diagnosis of RA, they had been treated with disease-modifying antirheumatic drugs (DMARDs) for at least 6 months, and they had a disease activity score (DAS) of at least 2.6.

**Interventions**

The study included a placebo group and a group receiving TRU-015. The TRU-015 group received a single intravenous infusion of TRU-015 at a dose of 10 mg/kg. The placebo group received a single intravenous infusion of placebo at the same time and dose.

**Measurements and Main Results**

The study found that TRU-015 was well-tolerated and that it significantly improved the clinical response in subjects with RA. The study also found that TRU-015 was effective in reducing the levels of anti-drug antibodies.

**Conclusions**

The study found that TRU-015 was well-tolerated and that it significantly improved the clinical response in subjects with RA. The study also found that TRU-015 was effective in reducing the levels of anti-drug antibodies.

**Limitations**

The study had several limitations, including a small sample size and a short duration. Further studies are needed to confirm the findings of this study.

**Future Studies**

Further studies are needed to evaluate the safety and efficacy of TRU-015 in larger groups of subjects with RA. These studies should also evaluate the long-term effects of TRU-015.

## Results

**Demographics (N=31)**

The study included 31 subjects with RA. The subjects were selected based on the following criteria: they had a confirmed diagnosis of RA, they had been treated with disease-modifying antirheumatic drugs (DMARDs) for at least 6 months, and they had a disease activity score (DAS) of at least 2.6.

**Adverse Events**

The study found that TRU-015 was well-tolerated and that it significantly improved the clinical response in subjects with RA. The study also found that TRU-015 was effective in reducing the levels of anti-drug antibodies.

**Serious Adverse Events**

The study found that there were no serious adverse events reported during the study. This suggests that TRU-015 is well-tolerated and that it is safe for use in subjects with RA.

**Adverse Events Occurring on Day of Infusion**

The study found that there were no adverse events reported on the day of infusion. This suggests that TRU-015 is well-tolerated and that it is safe for use in subjects with RA.

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## Conclusions

TRU-015 is a small modular immunopharmaceutical (SMIP) drug candidate that is designed to target B lymphocytes in RA. It is a CD20-directed therapy, which means it targets the CD20 protein on the surface of B lymphocytes. This protein is a key component of the B cell receptor, and its expression is increased in B lymphocytes in RA. By targeting CD20, TRU-015 is designed to deplete B lymphocytes and reduce the inflammation associated with RA.

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## Immunogenicity

TRU-015 is a small modular immunopharmaceutical (SMIP) drug candidate that is designed to target B lymphocytes in RA. It is a CD20-directed therapy, which means it targets the CD20 protein on the surface of B lymphocytes. This protein is a key component of the B cell receptor, and its expression is increased in B lymphocytes in RA. By targeting CD20, TRU-015 is designed to deplete B lymphocytes and reduce the inflammation associated with RA.

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## Conclusions

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